

Original Research Articles

Effects of different feeds and stocking densities on growth and survival rates of mud crab (*Scylla paramamosain*) at the stage from megalopa to crablet-1

Tien Hai Ly^{1a}, Le Hoang Vu^{2b}, Doan Xuan Diep^{3c}¹ Bac Lieu University, Bac Lieu City, Bac Lieu province, Vietnam, ² Faculty of Agriculture, Bac Lieu University, Bac Lieu City, Bac Lieu Province, Vietnam, ³ Van Hien University, Ho Chi Minh City, VietnamKeywords: mud crab, *Scylla paramamosain*, megalopa, feed, stocking density<https://doi.org/10.46989/001c.91128>

Israeli Journal of Aquaculture - Bamidgeh

Vol. 76, Issue 1, 2024

Mud crabs (*Scylla* genus) are luxury foods in high demand internationally. The efficient techniques for mud crab hatcheries are vital for providing breeds for their aquaculture, which is rapidly growing in many countries. This study aims to investigate the effects of different feeds and stocking densities on mud crabs' growth and survival rates (*Scylla paramamosain*) in the stage from megalopa to crablet-1 stage. Two separate experiments were conducted indoors in the 60-liter round plastic tanks (containing 50 liters of water at a 28‰ salinity). Experiment 1 investigated four feeds: frozen Artemia biomass, pureed shrimp meat, Lansy pellet feed (48% protein), and NRD pellet feed (55% protein). Megalopae (mean weight of 5.8 mg) were stocked at a density of 10/L. In experiment 2, the megalopae (mean weight of 5.4 mg) were stocked at densities of 20, 30, and 40/L and were fed the Lansy pellet feed, which was the best one selected from experiment 1. High survival rates were obtained at all four feeds (82.2–87.5%) and three stocking densities (88.4–90.1%). The growth performances in Lansy feed and frozen Artemia biomass were better than those in pureed shrimp meat and NRD pellet feed, which was seen through higher indicators of daily weight gain (DWG) and specific growth rate in weight (SGRw) ($p < 0.05$). Despite the survival rate showing no significant difference among the feeds, their highest value (87.5%) was observed in the Lansy pellet feed. On the other hand, the growth performances showed a significant decrease at stocking densities $\geq 30/L$, as complemented by a significant decrease in DWG and SGRw at these stocking densities ($p < 0.05$). Both survival rate and metamorphosis durations did not show significant differences among the feeds or stocking densities ($p > 0.05$). The investigated feeds and stocking densities suit the nursing mud crab (*S. paramamosain*) megalopa. In contrast, the Lansy pellet feeds had a stocking density of 20/L, resulting in the highest nursing efficiency.

INTRODUCTION

Mud crab is the common name of the crab species of the *Scylla* genus. Crabs are among the seafood items that are globally distributed, with a consistently growing demand.¹ It has been determined that four species are under the genus *Scylla*, including *Scylla olivacea*, *S. paramamosain*, *S. serrata*, and *S. tranquebarica*. All these species are known to have great potential for commercial aquaculture because they tolerate environments with poor water quality and are highly resistant to pathogens. Mud crabs are currently

cultured extensively in many countries, especially South-east Asian countries.¹⁻⁴ Among them, two species, *S. paramamosain* (locally called Lotus crab) and *S. olivacea* (called Fire crab), are distributed naturally in Vietnam, with over 95% of *S. paramamosain* and only about 5% of *S. olivacea* in their population.^{5,6}

S. paramamosain is recently considered one of the most important species for aquaculture in coastal areas of the Mekong Delta region of Vietnam, where the impacts of climate change and saltwater intrusion are more and more evident because it has good adaptability to environmental

a Equal contributor

b Equal contributor

c Corresponding author. E-mail: DiepDX@vhu.edu.vn

conditions in these areas and has high prices in both domestic and foreign markets.⁴ The Mekong Delta region has over 350,000 ha of culturing area for mud crabs, with a total production of about 38,000 tons⁴ out of around 53,000 tons produced in the country.⁷ Along with the development of commercial crab farming, the breed production industry of mud crabs has achieved snappy progress in the Mekong Delta, with over 600 hatcheries and 1.5 billion crablets produced annually in the region,⁸ providing almost all the breed for the farming. However, hatchery activities of mud crab still have limitations, even in Vietnam as well as many other nations, due to low survival rates, hence the lack of crablets for aquaculture on large scales.^{2,8,9}

The larval mud crabs often maintained in the hatcheries include three stages: zoea-1 to zoea-5 larvae, zoea-5 to megalopa, and megalopa to the crablet phase.¹⁰ The critical metamorphosis stage from megalopa to crablet-1 in mud crabs has often indicated inconsistent and unreliable results, with low survival rates.^{3,11} The mean survival rates of nursing mud crab megalopae often fluctuate from 5.0 to 58.0%.³ High mortality levels in this stage may be caused by: (i) nutrient factors (both quantity and quality) that result in the occurrence of Molting Death Syndrome¹²⁻¹⁴; (ii) cannibalism at high levels, as crabs in the megalopa stage possess pincers¹⁵⁻¹⁸; (iii) poor water quality¹⁹⁻²¹; and (iv) bacterial diseases, parasites, and fungi,^{22,23} among others. Therefore, the practice of directly stocking megalopa in ponds is not recommended. Instead, the nursery is needed to grow mud crabs of a larger size before pond stocking, and practical nursery techniques still need further development.²⁴

In the practice of nursing crustaceans, different feeds, like natural feeds, live feeds, and formulated feeds, have often been used in the stage of megalopa metamorphosis to the crablet stage. For the natural and live feeds, *Artemia* with live or frozen forms has often been practiced with the addition of prawn or shrimp meat, fish meat, and dried shrimp. Besides, *Acetes* shrimp,²⁵ dried mud worms, or artificial feed (larva-shrimp feed),^{3,9,10,26} minced trash fish, green mussels,¹⁵ brown mussels,¹ or artificial feed^{10,27-29} have also been used. Some previous studies have shown that the role of natural feed in the hatchery in the larval stages of mud crabs cannot be completely substituted for artificial feed, but the combination of them is still recommended.^{30,31} Still, in the megalopa stage, formulated particle feed could completely replace live prey.^{29,32} The development of pellet diets or aquafeed for aquaculture species has gained much interest nowadays, as pellets offer many advantages compared to natural feeds. In terms of nutrient content, artificial feed offers a nutritionally balanced diet with known nutrient content, such as total lipids and protein, that will promote growth and reproduction in the crustacean.³³ In the communal rearing technique, cannibalism is a major problem.²⁴ Manipulating the levels of protein and lipids is important for growth and reproduction in the crustacean group. The formulated feed could provide sufficient nutrition for them.³⁴ Our investigation aimed to compare mud crab's growth performance and survival rate (*S. paramamosain*) at the megalopa-to-crablet-1

stage nursed using different feeds (including pellets containing different protein contents, natural and live feeds) and different stocking densities. This is an important link for optimizing nursing techniques in the early stages of the crabs to develop commercial aquaculture of this crab species.

MATERIALS AND METHODS

EXPERIMENTAL MATERIALS

One-day-old megalopa of mud crabs (*S. paramamosain*), with a mean initial body weight of 5.40–5.80 mg and a mean initial carapace width of 0.65 mm, were used in the two experiments that were purchased at a hatchery in Bac Lieu City, Bac Lieu Province, Vietnam, and were transported in plastic bags at a density of 200–300/L.³⁵ After transport, megalopae were gradually acclimated for one week to the water salinity of 28‰ and temperature conditions of the experiments before they were stocked in experimental tanks.

Four feeds used in experiment 1 were included: (1) Frozen *Artemia* biomass (containing 54.5% protein, 12.9% total fat, and 11.2% moisture in dried biomass) was transferred from the farming area in Vinh Chau district, Soc Trang province, Vietnam, and stored cold (-21°C) for use throughout the experiment. Before feeding, *Artemia* was completely thawed and washed with fresh water to remove dirt in *Artemia* biomass; (2) pureed meat of Stork shrimp (*Metapenaeus tenuipes* Kubo, 1949), a locally common shrimp species that is often used as live feed in aquaculture. The average analyzed nutrient profile of shrimp meat includes protein 19.4±0.56%, lipid 1.15±0.19%, and water 76.3±0.57%³⁶; (3) Lansy pellet feed, a feed for shrimp post-larvae (containing contents of 48% protein, 9% fat, and 9% moisture); and (4) NRD pellet feed, a feed for ornamental fish (containing contents of protein: 55%, fat: 9%, and moisture: ≤ 8%). Both pellet feeds have a particle size of 500 µm and were made by INVE Company in Thailand. In experiment 2, Lansy pellet feed, which observed the highest crab performance in experiment 1, was used.

The water used for the two experiments was sediment-removed saline at 28‰ salinity. The water was treated with 30 mg/L chlorine and continuously strongly aerated for 3 to 4 days, then pumped into the experimental tanks through a filter bag with a mesh size of 5 µm.

The 60-liter round plastic tanks were used for the experiments.

The US National Research Council's guide for the Care and Use of Laboratory Animals was followed.

EXPERIMENTAL DESIGN

This study aimed to compare the effects of four feeds and three stocking densities on the growth and survival rates of mud crabs (*S. paramamosain*) at the stage from megalopa to crablet-1. The study included two separate experiments with a completely randomized design in three replications for each placed indoors. The experiments were carried out at the Aquaculture Experimental Hatchery of Bac Lieu Uni-



Figure 1. The nursery tanks were arranged with shelters

versity in South Vietnam from April to May 2020 and are described as follows:

EXPERIMENT 1: COMPARING THE GROWTH AND SURVIVAL RATES OF MUD CRABS (*S. PARAMAMOSAIN*) AT THE STAGE FROM MEGALOPA TO CRABLET-1 NURSED USING DIFFERENT FEEDS

This experiment aimed to assess the impact of four distinct feeds on mud crabs' growth and survival rates (*S. paramamosain*) during the transition from megalopa to crablet-1 stage. The experiment was conducted in twelve 60-liter round plastic tanks (containing 50 liters of water at a salinity of 28‰). The megalopae, with a mean weight of 5.80 mg and a mean carapace width of 0.65 mm, were stocked into the tanks at a density of 10/L. The crabs were fed four experimental feeds (frozen *Artemia* biomass, pureed shrimp meat, Lansy pellet feed, and NRD pellet feed) four times a day (6:00, 12:00, 18:00, and 24:00) according to a satisfying diet adjusted based on the situation of the nursery tank, fluctuating between 15-20% body weight per day. Plastic mesh panels with a mesh size of 4 mm were arranged into the bottoms of tanks as shelters for crabs, with a shelter density of 2 m² of plastic mesh per m² of tank bottom area³⁷ (Figure 1). The water in the tanks was constantly slightly aerated to ensure dissolved oxygen demand for the crabs during the nursing period. It was changed periodically every two days, changing 30% of the volume of the tanks with prepared water in the settling tanks. The fecal matter and uneaten feeds were removed daily from tanks at 21:00 by siphoning. Before siphoning, the aeration systems in the tanks were turned off to settle the sediment at the bottom of the tanks.

EXPERIMENT 2: COMPARING THE GROWTH AND SURVIVAL RATES OF MUD CRABS (*S. PARAMAMOSAIN*) AT THE STAGE FROM MEGALOPA TO CRABLET-1 NURSED AT DIFFERENT STOCKING DENSITIES

In this experiment, megalopae with a mean body weight of 5.4 mg and carapaces width of 0,65 mm were arranged into nine 60-litter-around plastic tanks (containing 50 liters of water at a salinity of 28‰) at stocking densities of 20, 30, and 40/L. The Lansy pellet feed, the best feed chosen from experiment 1, provided crabs with a daily diet to apparent satiation, equal to 15-20% of the biomass. The feed ratio was divided equally and given four times a day (6:00, 12:00,

18:00, and 24:00). The regimes of management and care in this experiment were done in the same manner as in experiment 1.

DATA COLLECTION

Water quality parameters in the tanks in both experiments were controlled before and during the nursing period, including temperature and pH, which were measured twice a day at 7:00 and 14:00, while TAN (N-NH₄⁺) and nitrite (N-NO₂⁻) were checked every three days at 14:00. All were tested using Sera GH test kits made in Germany.

Metamorphosis duration was determined from the megalopae stocked in the experiment tanks until they completely metamorphosed into crablet-1 through visual observation. The observation began after five days of stocking, with a frequency of every three hours.

Data for growth and survival rates were collected when the megalopae completely metamorphosed into crablet-1. The crablet-1 weights were weighed using an electrical analytical balance calibrated to 0.01 mg. In addition, ten s of crablet-1 from each tank were randomly collected and measured for the length and width of the carapace using biometric measurements under a microscope at a magnification of 10x with a millimeter unit. The monitored parameters of growth and survival rates from two experiments were determined using the following equations:

$DWG (g \text{ day}^{-1}) = (\text{final body weight} - \text{initial body weight}) / \text{duration of the experiment};$

$SGRw (\% \text{ day}^{-1}) = 100 (\ln(\text{final weight}) - \ln(\text{initial weight})) / \text{duration of experiment};$

$\text{Survival rate} (\%) = 100(\text{final number of crablets-1} / \text{initial number of megalopas})$

DATA ANALYSIS

The variance homogeneity was assessed using Levene's test. The percentage data were transformed to arcsine before conducting statistics. A one-way analysis of variance (ANOVA) together with Duncan's post-hoc tests were used to identify significant differences between the mean values (significance level of $p < 0.05$) using SPSS 20.0.

RESULTS

PARAMETERS OF NURSING PERFORMANCE USING DIFFERENT FEEDS

WATER QUALITY PARAMETERS

The water quality parameters monitored during the nursing period are presented in Table 1. The fluctuating ranges included temperature from 29.08 to 30.11 °C, pH from 7.8 to 8.00, TAN at 0.01 mg/L, and nitrite at 0.001 mg/L. Overall, they remained stable throughout the experiment.

GROWTH PERFORMANCE

Growth performance was determined based on parameters such as the mean weight, DWG, and SGRw, and the mean width and length of carapaces. Data in Table 2 shows that

Table 1. Water quality parameters during nursing mud crab (*S. paramamosain*) from megalopa to crablet-1 using different feeds

Feeds	Temperature (°C)		pH		TAN (mg/L)	NO ₂ ⁻ (mg/L)
	7:00	14:00	7:00	14:00		
Frozen Artemia biomass	29.1±0.2	30.1±0.2	7.8±0.1	7.8±0.1	0.01±0.00	0.001±0.000
Pureed shrimp meat	29.1±0.2	30.1±0.2	8.0±0.0	8.0±0.1	0.01±0.00	0.001±0.000
Lansy pellet feed	29.1±0.2	30.1±0.2	8.0±0.0	8.0±0.0	0.01±0.00	0.00±0.0000
NRD pellet feed	29.1±0.2	30.1±0.2	8.0±0.0	8.0±0.0	0.01±0.00	0.001±0.000

Table 2. Mean weight, width and length of carapace, DWG, and SGRw of mud crab (*S. paramamosain*) from megalopa to crablet-1 nursed using different feeds

Feeds	Frozen Artemia biomass	Pureed shrimp meat	Lansy pellet feed	NRD pellet feed
Initial weight (mg)	5.8 ±0.1	5.8 ±0.1	5.8 ±0.1	5.8 ±0.1
Final weight (mg)	9.3±0.2 ^a	8.8±0.1 ^b	9.4±0.1 ^a	9.0±0.2 ^{ab}
Initial carapace width (mm)	0.65±0.02	0.65±0.02	0.65±0.02	0.65±0.02
Final carapace width (mm)	2.96±0.16 ^a	2.94±0.17 ^a	2.97±0.10 ^a	2.94±0.10 ^a
Initial carapace length (mm)	3.80±0.22	3.80±0.22	3.80±0.22	3.80±0.22
Final carapace length (mm)	2.84±0.18 ^a	2.89±0.18 ^a	2.89±0.15 ^a	2.88±0.15 ^a
DWG (mg day ⁻¹)	0.61±0.02 ^a	0.52±0.01 ^b	0.62±0.01 ^a	0.53±0.01 ^b
SGRw(% day ⁻¹)	7.8±0.2 ^a	7.0±0.1 ^b	8.0±0.1 ^a	7.2±0.10 ^b

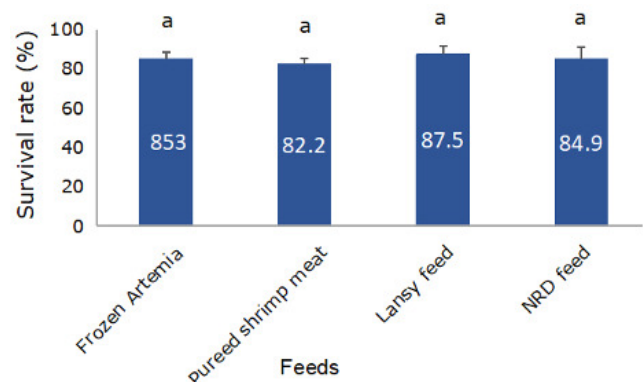
Values are presented as mean±SD. Values with different superscript letters (a, b) in the same rows show a significant difference ($p < 0.05$).

the final carapace width ranged from 2.94 to 2.97 mm and the final carapace length ranged from 2.84 to 2.89 mm; both showed no significant differences among the feeds used ($p > 0.05$). Meanwhile, the highest final weight was observed in Lansy shrimp pellet feed (9.4 mg), which did not show a significant difference from that in frozen Artemia biomass and NRD pellet feed (9.3 and 9.0 mg, respectively) ($p > 0.05$). The lowest final mean body weight was found in pureed shrimp meat (8.8 mg) and showed a significant difference from that in both Lansy pellet feed and frozen Artemia biomass ($p < 0.05$) but did not significantly differ from that in NRD pellet feed.

DWG and SGRw in Lansy feed and frozen Artemia biomass were significantly higher than those in pureed shrimp meat and NRD pellet feed ($p < 0.05$). Both did not show significant differences between frozen Artemia biomass (0.61 g day⁻¹ and 7.8% day⁻¹) and Lansy pellet feed (0.62 g day⁻¹ and 8.0% day⁻¹) or between pureed shrimp meat (0.52 g day⁻¹ and 7.0% day⁻¹) and NRD pellet feed (0.53 g day⁻¹ and 7.2% day⁻¹) (Table 2).

SURVIVAL RATE

The survival rates were observed to be rather high (82.2–87.5%) and did not show significant differences among the investigated feeds ($p > 0.05$). The highest survival rate was at Lansy pellet feed (87.5%), followed by frozen Artemia biomass (85.3%), NDR pellet feed (84.9%), and pureed shrimp meat (82.2%) (Figure 2).

**Figure 2. Survival rate of mud crab (*S. paramamosain*) from megalopa to crablet-1 nursed using different feeds**

[Values are presented as mean±SD. The bars with different letters (a, b) show a significant difference ($p < 0.05$)]

METAMORPHOSIS DURATION

Metamorphosis duration was shown to be relatively equivalent at feeds of frozen Artemia biomass, Lansy pellet feed, and NRD pellet feed (10.03 days). Meanwhile, a longer metamorphosis duration was observed in pureed shrimp meat (11.67 days). Here, there were no significant differences in metamorphosis durations among feeds used ($p > 0.05$) (Figure 3).

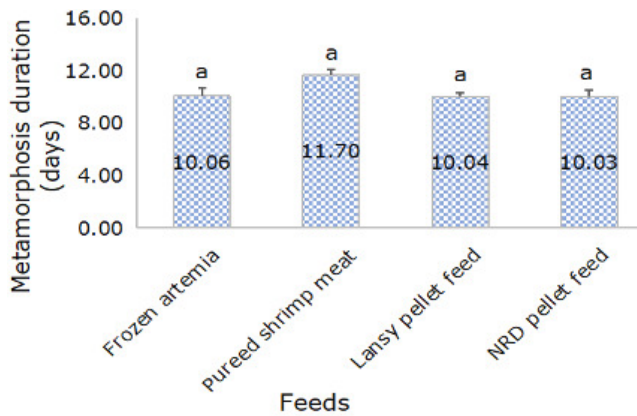


Figure 3. Metamorphosis duration of mud crab (*S. paramamosain*) from megalopa to crablet-1 nursed using different feeds

[Values are presented as mean±SD. The bars with different letters (a, b) show a significant difference ($p < 0.05$)]

PARAMETERS OF NURSING PERFORMANCE AT DIFFERENT STOCKING DENSITIES

WATER QUALITY PARAMETERS

In this experiment, water quality parameters were observed: temperature fluctuated between 27.7 and 29.6 °C; pH 7.7 and 7.9; TAN 0.26 and 0.31 mg/L; and nitrite at a concentration of 0.01 mg/L (Table 3).

GROWTH PERFORMANCE

Despite final carapace width (ranging from 2.99–3.33 mm) showing no difference among stocking densities, final weight (9.3–9.9 mg), the final carapace lengths (2.90–2.95 mm), DWG (0.66–0.76 mg day⁻¹), and SGRw (9.1–10.2% day⁻¹) were shown to decrease with increases in the stocking densities, and significant decreases were at a stocking density of 40/L ($p < 0.05$). For DWG and SGRw, significant reductions were shown at stocking densities ≥ 30 /L (Table 4).

SURVIVAL RATE

The survival rates observed were pretty high and relatively equal among stocking densities, with values of 89.7%, 90.1%, and 88.4% at stocking densities of 20, 30, and 40/L, respectively (Figure 4).

Table 3. Water quality parameters during nursing mud crab (*S. paramamosain*) from megalopa to crablet-1 at different stocking densities

Densities	Temperature (°C)		pH		TAN (mg/L)	NO ₂ ⁻ (mg/L)
	7:00	14:00	7:00	14:00		
20	27.8±0.1	29.4±1.1	7.9±0.2	7.9±0.2	0.26±0.05	0.001±0.000
30	27.8±0.1	29.3±1.2	7.7±0.2	7.7±0.2	0.31±0.09	0.00±0.0000
40	27.7±0.6	29.6±1.2	7.8±0.2	7.8±0.2	0.29±0.08	0.001±0.000

METAMORPHOSIS DURATION

Figure 5 shows that metamorphosis durations from megalopa to crablet-1 had little variation among stocking densities of 20–40/L. They were observed ranging from 12.01 to 12.03 days, and the difference was not found among stocking densities.

DISCUSSION

WATER QUALITY PARAMETERS

Both experiments were conducted indoors, so the water quality parameters showed little fluctuation. The temperature ranged from 29.08 to 30.11 °C, pH from 7.8 to 8.00, TAN at 0.01 mg/L, and nitrite at 0.001 mg/L in experiment 1 (Table 1), and they were 27.7–29.6 °C, 7.7–7.9; 0.26–0.31 mg/L; and 0.001 mg/L, respectively, in experiment 2 (Table 3). Overall, all of them sustained the proper conditions for the life of *Scylla* crabs during the nursing period. These water quality parameters suggested for mud crab rearing in larval and adult stages were a temperature of 28–30 °C (Ye et al., 2011; Omn³), a pH of 7.5–8.5 (0.5 daily variations),³⁸ TAN < 3 mg/L,³⁹ and NO₂⁻ < 10 mg/L at salinity >15‰.³⁹ Furthermore, it was concluded that the water quality parameters for a shrimp culture can be used as a guide in mud crab farming activities.^{24,40} Probably because the water in the tanks was changed periodically and the tanks were siphoned daily, the nitrite concentration was low during larval rearing in all experiment tanks.

PERFORMANCE OF THE CRABS NURSED USING DIFFERENT FEEDS

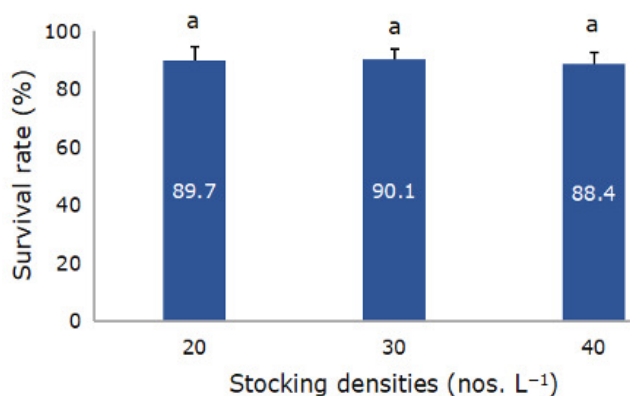
The gastrointestinal tracts and articulated appendages of larval crustaceans show remarkable morphological ranges, enabling them to capture and process different feed sources. Therefore, the diet of larval crustaceans frequently changes markedly over developmental stages, depending on their size and feeding abilities.⁴¹ Crab larvae, as well as homarids and carideans, do not have the anterior midgut diverticula and alternative sources of digestive enzymes as those of Penaeid shrimp larvae. Therefore, the choice of appropriate feed types for the rearing of mud crab larvae, especially at the stages of zoea-5 and megalopa, is very important for the yields.⁴²

Experiment 1 showed the highest growth performance with Lansy pellet feed and frozen Artemia biomass. This was evident as the highest values of final weight, DWG,

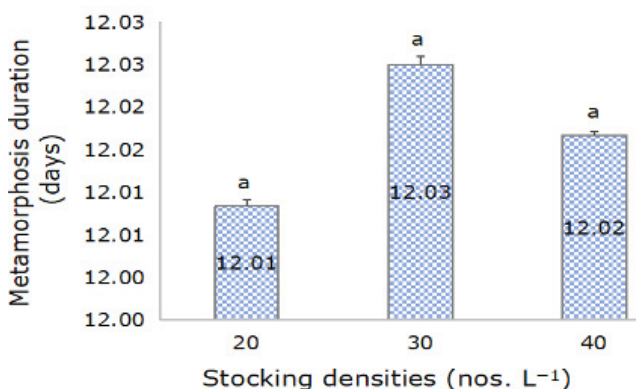
Table 4. Mean body weight, width and length of carapace, DWG, and SGRw of mud crab (*S. paramamosain*) from megalopa to crablet-1 nursed at different stocking densities

Densities (/L)	20	30	40
Initial weight (mg)	5.4 ± 0.7	5.4 ± 0.7	5.4 ± 0.7
Final weight (mg)	9.9 ± 0.1 ^a	9.7 ± 0.1 ^{ab}	9.3 ± 0.1 ^b
Initial carapace width (mm)	0.65 ± 0.02	0.65 ± 0.02	0.65 ± 0.02
Final carapace width (mm)	3.00 ± 0.08 ^a	3.33 ± 3.16 ^a	2.99 ± 0.07 ^a
Initial carapace length (mm)	3.80 ± 0.22	3.80 ± 0.22	3.80 ± 0.22
Final carapace length (mm)	2.95 ± 0.11 ^a	2.94 ± 0.11 ^{ab}	2.90 ± 0.17 ^b
DWG (mg day ⁻¹)	0.76 ± 0.01 ^a	0.70 ± 0.01 ^b	0.66 ± 0.01 ^c
SGRw (%) day ⁻¹)	10.2 ± 0.1 ^a	9.6 ± 0.2 ^b	9.1 ± 0.6 ^c

Values are presented as mean ± SD. Values with different superscript letters (a, b) in the same rows show a significant difference ($p < 0.05$).

**Figure 4. Survival rate of mud crab (*S. paramamosain*) from megalopa to crablet-1 nursed at different stocking densities**

[Values are presented as mean ± SD. The bars with different letters (a, b) show a significant difference ($p < 0.05$)]

**Figure 5. Metamorphosis duration of mud crab (*S. paramamosain*) from megalopa to crablet-1 nursed at different stocking densities**

[Values are presented as mean ± SD. The bars with different letters (a, b) show a significant difference ($p < 0.05$)]

and SGRw were observed in these feeds. Meanwhile, the lowest growth performance was with the pureed shrimp

meat (Table 2). Differences in the nutritional composition of the investigated feeds could cause the differences in crab performance observed. Three of the four investigated feeds—the Lansy pellet feed, frozen Artemia biomass, and NRD pellet feed—contained between 48% and 55% protein and 9% and 12.9% lipid, which are within levels suggested for the growth of mud crabs.^{43–45} Namely, the best growth performance and nutrient turnover of mud crab (*S. serrata*) juveniles were found in a 45% crude protein diet.⁴⁵ Mud crabs (*S. serrata*) grew well when they were fed with 32–40% dietary protein and 6–12% lipids.⁴⁴ Dietary lipid levels ranging from 5.3% to 13.8% appeared to meet the lipid requirement of juvenile mud crab *S. serrata*.⁴³ Meanwhile, the pureed meat of Stork shrimp contains a lower protein and lipid content, with only 19.4 ± 0.56% and 1.15 ± 0.19%, respectively. This might be the cause of the lower weight gain of crabs consuming this feed. The growth performance of crablet-1 of *S. paramamosain* crabs fed fresh shrimp meat was lower than that of frozen Artemia biomass, which was also found by Anh et al.⁴⁶ In addition, the growth performance of *S. paramamosain* megalopa reached its highest level with Lansy pellet feed (containing 48% protein) and decreased with NRD pellet feed (containing 55% protein), which was consistent with the previous study on *S. serrata* juvenile.⁴⁵ The researchers highlighted that higher protein levels in the diets of animals compared to their protein nutrient requirements could lead to an imbalance in the dietary nutrient profile and result in excess protein catabolism. This could generate higher ammonia contents in their hemolymph as a product of excretion.⁴⁷ Accumulation of ammoniac nitrogen in the hemolymph could negatively affect metabolic processes such as transporting oxygen and balancing osmotic pressure.^{48,49} It can increase the metabolic cost of nitrogen excretion.^{50,51} In addition, at excessively high dietary protein levels, free amino acids accumulated in the body fluids may become toxic, affecting normal metabolism and compromising normal growth in crustaceans.⁵² The information mentioned above may explain the low growth performance of experimental crabs fed NRD pellet feed compared to Lansy pellet feed. Besides, it recommended that in the practice of nursing crustaceans in the megalopa stage, formulated particle feed could completely replace live prey,^{29,32} as pellets offer many advan-

tages compared to natural or live feeds. In terms of nutrient content, artificial feed offers a nutritionally balanced diet with known nutrient content, such as total lipids and protein, that will promote growth and reproduction in the crustacean.^{33,34} Results of our study also showed that, although the difference was not significant, Lansy pellets resulted in crab performance slightly higher than frozen *Artemia* biomass feed.

Despite the survival rates and metamorphosis durations showing no significant difference among the investigated feeds, the highest survival rate was observed in the Lansy pellet feed (Figure 2). Meanwhile, the lowest survival rate (Figure 2) and the longest metamorphosis duration (Figure 3) were observed for pureed shrimp meat. It is suggested that feed availability (quantity and quality) is one of the major factors influencing survival in reared crabs.^{53,54} The high mortality during the molting phases is often attributed to the poor nutritional status of the larvae.⁹ Besides, the shortage of suitable feed is one of the factors leading to cannibalism and decreasing survival in crabs.⁵⁵⁻⁵⁸ Survival rates (82.2–87.5%) in *S. paramamosain megalopa* in the current study were higher as compared to some other species of the *Sylla* genus recorded in previous studies. *S. serrata* had survival rates of 38, 46, and 48% with feeds of nauplii *Artemia*, enriched nauplii *Artemia*, and frozen crab larvae, respectively,⁹ 32.8±4.8% with feeds of mussels or small shrimps (*Acetes sp.*),⁵⁵ and 48.3–53.3% with feeds of macerated brown mussel meat (*Modiolus metcalfei*) or fish.¹⁵ *Sylla sp.* obtained a 30–50% survival rate with feeds of newly hatched and adult *Artemia*.⁵⁹ *S. paramamosain* had the highest survival rate of 72.9% when fed 100% Lansy-shrimp feed (called Lansy pellet feed in the present study).²⁹

In addition, the metamorphosis duration in the megalopa-to-crablet-1 stage of *S. paramamosain* (10.03–11.70 days) (Figure 3) was within the ranges recorded in species of the *Sylla* genus in other studies. The megalopa mud crab stage often lasts 7–10 days before molting to the first crab stage.² Mud crab megalopae took 11–12 days to reach the first crab stage at a salinity of 31‰, while it took only 7–8 days at a lower salinity of 24–31‰.⁶⁰ The megalopa duration was recorded as unusually long (15–16 days) at 29°C and a salinity of 34–35‰.⁶¹ These findings indicated that the four feeds were highly effective for nursing the mud crab *S. paramamosain* at the megalopa to crablet-1 stage. The best feeds were Lansy pellet, frozen *Artemia* biomass, NRD pellet feed, and pureed shrimp meat. Besides, the results also show that NRD pellet feed (55%) decreased the performance of the experiment crabs.

PERFORMANCE OF THE CRABS NURSED AT DIFFERENT DENSITIES

The increases in stocking density may significantly improve production yield per unit area. However, too high a stocking density of crab megalopa can also lead to competition for space and feed utility⁶² and crowd physiological stress.⁶³ Also, the cannibalistic nature of megalopa with claws and free swimming causes sub-lethal injuries (e.g., limb autotomy),⁶⁴ reducing the energy available for growth,⁶⁴ and

affecting the health, growth, and survival of farmed animals.⁶⁵⁻⁶⁷ The present study indicated the change in stocking densities from 20 to 40/L had no clear effect on the survival rate and metamorphosis duration of *S. paramamosain* from megalopa to crablet-1 (Figures 4 and 5). Meanwhile, growth performance significantly decreased with a stocking density ≥ 30/L, with daily weight gain and SGR in weight significantly lower at these stocking densities ($p < 0.05$) (Table 4).

The survival rates (88.4–90.1%) in *S. paramamosain* in this experiment were higher than those found in several previous studies on mud crabs in the same stage under different conditions of nursing.^{9,10,15,29,35,55,59,68-70} It has been endorsed that in the communal rearing technique of crabs, cannibalism is a critical problem.^{24,29} The adequate availability of feed and shelter could reduce the rate of cannibalism.³⁴ In this experiment, we used Lansy pellet feed, which resulted in the best growth and survival rates for the crabs from experiment 1, while simultaneously putting plastic mesh panels in the tanks, providing sufficient shelter to the crabs. Also, the experimental tanks were placed indoors, changed periodically, and siphoned waste every day, so the water in the tanks was maintained at good quality with few fluctuations, perhaps helping improve the crabs' survival rate. Furthermore, Lansy pellet feed with stocking densities of 20–40/L has not had a remarkable variation in the metamorphosis duration of mud crab *S. paramamosain* in the reared stage from megalopa to crablet-1. The metamorphosis duration (12.01–12.03 days) in this experiment was within the range found in previous studies in some species of mud crabs.^{2,60,61} However, stocking densities ≥ 30/L significantly reduced the growth performance in DWG and SGRw (Table 4). It is recommended that Lansy feed be commonly used for rearing crabs from megalopa to crablet-1, and stocking density should be applied at 20/L to achieve the highest rearing efficiency in our conditions.

CONCLUSION

Four investigated feed types all showed great potential in nursing mud crab (*S. paramamosain*) at the stage from megalopa to crablet-1, with high survival rates in crablet-1 (82.2–90.1%). Among these the Lansy pellet feed (containing 48% protein) and stocking density of 20 /L offered the highest nursing efficiency within the indoor nursing system in the tanks. Also, NRD pellet feed (containing 55% protein) showed a decrease in crab performance. The findings have provided valuable information for choosing appropriate feeds and stocking densities to reduce costs and enhance nursing performance in *S. paramamosain* megalopa. Further studies need to be conducted, including the use of Lansy pellet feed for nursing *S. paramamosain* megalopa at higher stocking densities with nursing systems of hapas and ponds or applying biofloc nursing systems to improve the efficiency of the nursing processes increasingly.

AUTHORS' CONTRIBUTION USING [CREDIT](#)

Conceptualization: Tien Hai Ly (Equal), Le Hoang Vu (Equal), Doan Xuan Diep (Equal). Methodology: Tien Hai Ly (Equal), Doan Xuan Diep (Equal). Writing – original draft: Tien Hai Ly (Lead). Writing – review & editing: Le Hoang

Vu (Equal), Doan Xuan Diep (Equal). Formal Analysis: Doan Xuan Diep (Lead). Investigation: Doan Xuan Diep (Lead).

Submitted: September 29, 2023 CST, Accepted: November 25, 2023 CST



This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CCBY-NC-ND-4.0). View this license's legal deed at <https://creativecommons.org/licenses/by-nc-nd/4.0> and legal code at <https://creativecommons.org/licenses/by-nc-nd/4.0/legalcode> for more information.

REFERENCES

1. Gunarto ., Herlinah . Level of crablet production in mangrove crab *Scylla paramamosain* with feeding enrichment using HUFA and vitamin C on larvae stages. *J Ilmu dan Teknologi Kelautan Tropis*. 2016;7(2):511-520. [doi:10.29244/jitkt.v7i2.10997](https://doi.org/10.29244/jitkt.v7i2.10997)
2. Waiho K, Fazhan H, Quinitio ET, et al. Larval rearing of mud crab (*Scylla*): What lies ahead. *Aquaculture*. 2018;493:37-50. [doi:10.1016/j.aquaculture.2018.04.047](https://doi.org/10.1016/j.aquaculture.2018.04.047)
3. Omn KK. Current practices in juvenile mud crab rearing. *AQUA Culture Asia Pacific Magazine*. 2013;9(4):44-46.
4. Hai TN. *Principles and Technology of Mud Crab Culture*. Agriculture Publishing House, Ha Noi; 2017.
5. Keenan C, Davie PJ, Mann DL. A revision of the genus *Scylla* de Haan, 1833 (Crustacea: Decapoda: Brachyura: portunidae). *The Raffles Bulletin of Zoology*. 1998;46(1):217-245.
6. Vay LL. Ecology and management of mud crab *Scylla* spp. Proceedings of the International forum on the culture of Portunid crabs. *Asian Fisheries Science*. 2001;14(2):101-111. [doi:10.33997/j.afs.2001.14.2.001](https://doi.org/10.33997/j.afs.2001.14.2.001)
7. Hungria DB, dos Santos Tavares CP, Pereira LÂ, Silva UAT, Ostrensky A. Global status of production and commercialization of soft-shell crabs. *Aquacult Int*. 2017;25(6):2213-2226. [doi:10.1007/s10499-017-0183-5](https://doi.org/10.1007/s10499-017-0183-5)
8. Hai TN, Vinh PQ, Việt LQ. Technical and financial aspects of mud crab hatcheries in the Mekong Delta. *Can Tho University Journal of Sciences*. 2018;54(1):169-175. [doi:10.22144/ctu.jsi.2018.022](https://doi.org/10.22144/ctu.jsi.2018.022)
9. Williams GR, Wood J, Dalliston B, Shelley CC, Kuo CM. *Mud Crab (Scylla Serrata) Megalopa Larvae Exhibit High Survival Rates on Artemia-Based Diets*. *Mud Crab Aquaculture and Biology*. ACIAR Proceeding; 1999.
10. Syafaat MN, Gunarto, Usman. The rearing of megalopa mud crab (*Scylla serrata*) using different formulated feed as additional source of food. In: *Proceeding of Forum Inovasi Teknologi Akuakultur. Conference: Asian Pacific Aquaculture*. Surabaya; 2016:209-214.
11. Mann D, Paterson B. Status of crab seed production and grow-out in Queensland. *Mud Crab Aquaculture in Australia and Southeast Asia*. In: *Proceedings of the ACIAR Crab Aquaculture Scoping Study and Workshop*. Australian Centre for International Agricultural Research; 2003:36-41.
12. Castine S, Southgate PC, Zeng C. Evaluation of four dietary protein sources for use in microbound diets fed to megalopae of the blue swimmer crab, *Portunus pelagicus*. *Aquaculture*. 2008;281(1-4):95-99. [doi:10.1016/j.aquaculture.2008.06.008](https://doi.org/10.1016/j.aquaculture.2008.06.008)
13. Hassan A, Hai TN, Chatterji A, Sukumaran N. Preliminary study on the feeding regime of laboratory reared mud crab larva, *Scylla serrata* (Forsskal, 1775). *World Applied Sciences Journal*. 2011;14(11):1651-1654.
14. Ikhwanuddi M, Azra MN, Sung YY, Munafi ABA, Shabdin ML. Live foods for juveniles' production of blue swimming crab, *Portunus pelagicus* (Linnaeus, 1766). *J of Fisheries and Aquatic Science*. 2012;7(4):266-278. [doi:10.3923/jfas.2012.266.278](https://doi.org/10.3923/jfas.2012.266.278)
15. Rodríguez EM, Quinitio FD, Parado-Estepa FD, Millamena OM. Culture of *Scylla serrata* megalops in brackishwater ponds. *Asian Fisheries Science*. 2001;14(2):185-189. [doi:10.33997/j.afs.2001.14.2.008](https://doi.org/10.33997/j.afs.2001.14.2.008)
16. Marshall S, Warburton K, Paterson B, Mann D. Cannibalism in juvenile blue-swimmer crabs *Portunus pelagicus* (Linnaeus, 1766): effects of body size, moult stage and refuge availability. *Applied Animal Behaviour Science*. 2005;90(1):65-82. [doi:10.1016/j.applanim.2004.07.007](https://doi.org/10.1016/j.applanim.2004.07.007)
17. Ut VN, Le Vay L, Nghia TT, Hanh TT. Development of nursery culture techniques for the mud crab *Scylla paramamosain* (Estampador). *Aquaculture Res*. 2007;38(14):1563-1568. [doi:10.1111/j.1365-2109.2006.01608.x](https://doi.org/10.1111/j.1365-2109.2006.01608.x)
18. Ye H, Tao Y, Wang G, Lin Q, Chen X, Li S. Experimental nursery culture of the mud crab *Scylla paramamosain* (Estampador) in China. *Aquacult Int*. 2010;19(2):313-321. [doi:10.1007/s10499-010-9399-3](https://doi.org/10.1007/s10499-010-9399-3)
19. Bryars SR, Havenhand JN. Effects of constant and varying temperatures on the development of blue swimmer crab (*Portunus pelagicus*) larvae: Laboratory observations and field predictions for temperate coastal waters. *Journal of Experimental Marine Biology and Ecology*. 2006;329(2):218-229. [doi:10.1016/j.jembe.2005.09.004](https://doi.org/10.1016/j.jembe.2005.09.004)

20. Romano N, Zeng C. The effects of salinity on the survival, growth and haemolymph osmolality of early juvenile blue swimmer crabs, *Portunus pelagicus*. *Aquaculture*. 2006;260(1-4):151-162. doi:10.1016/j.aquaculture.2006.06.019
21. Liao YY, Wang HH, Lin ZG. Effect of ammonia and nitrite on vigour, survival rate, moulting rate of the blue swimming crab *Portunus pelagicus* zoea. *Aquacult Int*. 2011;19(2):339-350. doi:10.1007/s10499-010-9398-4
22. Morado JF. Protistan diseases of commercially important crabs: A review. *Journal of Invertebrate Pathology*. 2011;106(1):27-53. doi:10.1016/j.jip.2010.09.014
23. Wang W. Bacterial diseases of crabs: A review. *Journal of Invertebrate Pathology*. 2011;106(1):27-53. doi:10.1016/j.jip.2010.09.018
24. Syafaat MN, Azra MN, Waiho K, et al. A review of the nursery culture of mud crabs, genus *Scylla*: Current progress and future directions. *Animals*. 2021;11(7):2034. doi:10.3390/ani11072034
25. Macintosh DJ, Goncalves F, Soares AMVM, Moser SM, Paphavisit N. Transport mechanisms of crab megalopae in mangrove ecosystems, with special reference to a mangrove estuary in Ranong, Thailand. In: *Mud Crab Aquaculture and Biology, ACIAR Proceeding*. ; 1999:178-186. <https://www.aciar.gov.au/file/68781/download?token=NhupYsCY>
26. Usman U, Kamaruddin K, Laining A. Substitusi penggunaan nauplius *Artemia* dengan pakan mikro dalam pemeliharaan larva kepiting bakau, *Scylla olivacea*. *Jurnal Riset Akuakultur*. 2018;13(1):29. doi:10.15578/jra.13.1.2018.29-38
27. Holme MH, Zeng C, Southgate PC. Use of microbound diets for larval culture of the mud crab, *Scylla serrata*. *Aquaculture*. 2006;257(1-4):482-490. doi:10.1016/j.aquaculture.2006.03.014
28. Holme MH, Zeng C, Southgate PC. The effects of supplemental dietary cholesterol on growth, development and survival of mud crab, *Scylla serrata*, megalopa fed semi-purified diets. *Aquaculture*. 2006;261(4):1328-1334. doi:10.1016/j.aquaculture.2006.08.032
29. Nguyen TP, Nguyen TE, Nguyen TKH, Le QV, Do TTH. Effects of different temperatures on the growth and survival of mud crab (*Scylla paramamosain*) larvae. *CTUJS*. 2021;13(3):9-16. doi:10.22144/ctu.jen.2021.035
30. Mardjono M, Hamid AN, Djunaidah IS, Satyantini WH. *Manual for Seed Production of Mud Crab (Scylla Serrata)*. Brackishwater Aquaculture Development Center; 1994.
31. Quinito ET, Parado-Esteva F, Alava V. Development of hatchery techniques for the mud crab *Scylla serrata*: 1. Comparison of different feeding schemes. *Mud Crab Aquaculture and Biology*. In: *Proceedings of an International Scientific Forum Held in Darwin, Australia. 21-24 April, 1997. ACIAR Proceedings*, 78. ACIAR Publishing; 1999:125-130.
32. Holme MH, Southgate PC, Zeng C. Survival, development and growth response of mud crab, *Scylla serrata*, megalopae fed semi-purified diets containing various fish oil:corn oil ratios. *Aquaculture*. 2007;269(1-4):427-435. doi:10.1016/j.aquaculture.2007.05.024
33. Magondu EW, Charo-Karisa H, Verdegem MCJ. Effect of C/N ratio levels and stocking density of *Labeo victorianus* on pond environmental quality using maize flour as a carbon source. *Aquaculture*. 2013;410-411:157-163. doi:10.1016/j.aquaculture.2013.06.021
34. Lee DOC, Wickins JF. *Crustacean Farming*. Blackwell Scientific Publications, Oxford; 1992.
35. Quinito ET, Parado-Esteva FD. Transport of *Scylla serrata* megalopa at various densities and durations. *Aquaculture*. 2000;185:63-71. doi:10.1016/S0044-
36. Dayal JS, Ponniah AG, Khan HI, Babu EPM, Ambasankar K, Vasagam KPK. Shrimps - a nutritional perspective. *Current Science*. 2013;104(11):1487-1491.
37. Việt LQ, Hải TN. Effects of water level, stocking density, and emerged substrate on survival rate of crab (*Scylla paramamosain*) larva from megalop stage to crab 1. *Can Tho University, Journal of Science*. 2018;54(3)(3):132. doi:10.22144/ctu.jvn.2018.049
38. Rodriguez EM, Parado-Esteva FD, Quinito ET. Extension of nursery culture of *Scylla serrata* (Forsskål) juveniles in net cages and ponds. *Aquaculture Res*. 2007;38(14):1588-1592. doi:10.1111/j.1365-2109.2007.01725.x
39. Ganesh K, Raj YCTS, Perumal S, Srinivasan P, Sethuramalingam A. Breeding, larval rearing and farming of mangrove crab, *Scylla serrata* (Forsk., 1775). *Advances in Marine and Brackishwater Aquaculture*. Published online 2015:163-172. doi:10.1007/978-81-322-2271-2_14
40. Shelley C, Lovatelli A. *Mud Crab Aquaculture—A Practical Manual*. *FAO Fisheries and Aquaculture Technical Paper*. 2011;(567):78.

41. Andrew J, O'Rourke R, eds. Feeding and nutrition of crustacean larvae. In: *Developmental Biology and Larval Ecology: The Natural History of the Crustacea*. Vol 7. Oxford University Press; 2020:310-333. doi:10.1093/oso/9780190648954.001.0001
42. Serrano, Jr. A. E. Changes in gut evacuation time of larval mud crab, *Scylla serrata* (Crustacea: Portunidae) fed artificial plankton or live food. *AACL Bioflux*. 2012;5(4):240-248.
43. Sheen SS, Wu SW. The effects of dietary lipid levels on the growth response of juvenile mud crab *Scylla serrata*. *Aquaculture*. 1999;175(1-2):143-153. doi:10.1016/s0044-8486(99)00027-7
44. Catacutan MR. Growth and body composition of juvenile mud crab, *Scylla serrata*, fed different dietary protein and lipid levels and protein to energy ratios. *Aquaculture*. 2002;208(1-2):113-123. doi:10.1016/s0044-8486(01)00709-8
45. Unnikrishnan U, Paulraj R. Dietary protein requirement of giant mud crab *Scylla serrata* juveniles fed iso-energetic formulated diets having graded protein levels. *Aquaculture Research*. 2010;41(2):278-294. doi:10.1111/j.1365-2109.2009.02330.x
46. Anh NTN, Ut VN, Wille M, Hoa NV, Sorgeloos P. Effect of different forms of *Artemia* biomass as a food source on survival, molting and growth rate of mud crab (*Scylla paramamosain*). *Aquaculture Nutrition*. 2010;17(2):e549-e558. doi:10.1111/j.1365-2095.2010.00796.x
47. Rosas C, Sanchez A, Díaz E, et al. Oxygen consumption and ammonia excretion of *Penaeus setiferus*, *P. schmitti*, *P. duorarum* and *P. notialis* postlarvae fed purified test diets: effect of protein level on substrate metabolism. *Aquat Living Resour*. 1995;8(2):161-169. doi:10.1051/alr:1995013
48. Schmitt ASC, Santos EA. Haemolymph nitrogenous constituents and nitrogen efflux rates of juvenile shrimp, *Penaeus paulensis* (Perez-Farfante), exposed to ambient ammonia-N. *Aquaculture Research*. 1999;30(1):1-11. doi:10.1046/j.1365-2109.1999.00268.x
49. Guzman C, Gaxiola G, Rosas C, Torre-blnco A. The effect of dietary protein and total energy content on digestive enzyme activities, growth and survival of *Litopenaeus setiferus* (Linnaeus 1767) postlarvae. *Aquaculture Nutrition*. 2001;7(2):113-122. doi:10.1046/j.1365-2095.2001.00161.x
50. Jauncey K. The effects of varying dietary protein level on the growth, food conversion, protein utilization and body composition of juvenile tilapias (*Sarotherodon mossambicus*). *Aquaculture*. 1982;27(1):43-54. doi:10.1016/0044-8486(82)90108-9
51. Vergara JM, Fernandez-Palacios H, Robaina L, Jauncey K, De La Higuera M, Izquierdo M. The effects of varying dietary protein level on the growth, feed efficiency, protein utilization and body composition of gilthead sea bream fry. *Fisheries science*. 1996;62(4):620-623. doi:10.2331/fishsci.62.620
52. Harper AE, Benevenga NJ, Wohlhueter RM. Effects of ingestion of disproportionate amounts of amino acids. *Physiological Reviews*. 1970;50(3):428-558. doi:10.1152/physrev.1970.50.3.428
53. Hartnoll RG. *Growth. The Biology of Crustacea*. Academic; 1982.
54. Sheen SS. Dietary cholesterol requirement of juvenile mud crab *Scylla serrata*. *Aquaculture*. 2000;189(3-4):277-285. doi:10.1016/s0044-8486(00)0379-3
55. Qunitio ET, Parado-Esteba FD, Millamena OM, Rodriguez E, Borlongan E. Seed production of mud crab *Scylla serrata* juveniles. *Asian Fisheries Science*. 2001;14(2):161-174. doi:10.33997/j.afs.2001.14.2.006
56. Genodepa J, Zeng C, Southgate PC. Preliminary assessment of a microbound diet as an *Artemia* replacement for mud crab, *Scylla serrata*, megalopa. *Aquaculture*. 2004;236(1-4):497-509. doi:10.1016/j.aquaculture.2004
57. Møller H, Lee SY, Paterson B, Mann D. Cannibalism contributes significantly to the diet of cultured sand crabs, *Portunus pelagicus* (L.): A dual stable isotope study. *Journal of Experimental Marine Biology and Ecology*. 2008;361(2):75-82. doi:10.1016/j.jembe.2008.04.013
58. Roslan S, Taher S, Ehteshamei F, Arshad A, Romano N. Effects of dietary peppermint (*Mentha piperita*) essential oil on survival, growth, cannibalism and hepatopancreatic histopathology of *Portunus pelagicus* juveniles. *Journal Environmental Biology*. 2016;37(4):785-790.
59. Qunitio ET, Parado-Esteba FD, Rodriguez E. Seed production of mud crab *Scylla* spp. *Aquaculture Asia*. 2002;7(3):29-31. http://hdl.handle.net/10862/3117
60. Ong KS. The early developmental stages of *Scylla serrata* Forskal reared in the laboratory. *Proceedings of Indo-Pacific Fisheries Council*. 1964;11:135-146.
61. Hamasaki K. Effects of temperature on the egg incubation period, survival and development period of the larvae of the mud crab *Scylla serrata* (Forsskl) (Brachyura: Portunidae) reared in the laboratory. *Aquaculture*. 2003;219(1-4):561-572. doi:10.1016/s0044-8486(02)00662-2

62. Refstie T, Kittelsen A. Effect of density on growth and survival of artificially reared Atlantic salmon. *Aquaculture*. 1976;8(4):319-326. [doi:10.1016/0044-8486\(76\)90114-9](https://doi.org/10.1016/0044-8486(76)90114-9)
63. Efrizal E. Effects of stocking density on survival rate and larval development of blue swimming crab, *Portunus pelagicus* (Linnaeus, 1758) under laboratory conditions. *AACL Bioflux*. 2017;10(2):217-226.
64. Sainte-Marie B, Lafrance M. Growth and survival of recently settled snow crab *Chionoecetes opilio* in relation to intra- and intercohort competition and cannibalism: A laboratory study. *Mar Ecol Prog Ser*. 2002;244:191-203. [doi:10.3354/meps244191](https://doi.org/10.3354/meps244191)
65. Rowland SJ, Mifsud C, Nixon M, Boyd P. Effects of stocking density on the performance of the Australian freshwater silver perch (*Bidyanus bidyanus*) in hapas. *Aquaculture*. 2006;253(1-4):301-308. [doi:10.1016/j.aquaculture.2005.04.049](https://doi.org/10.1016/j.aquaculture.2005.04.049)
66. Islam MS, Rahman MM, Tanaka M. Stocking density positively influences the yield and farm profitability in cage aquaculture of sutchi catfish, *Pangasius sutchi*. *J Appl Ichthyol*. 2006;22(5):441-445. [doi:10.1111/j.1439-0426.2006.00746.x](https://doi.org/10.1111/j.1439-0426.2006.00746.x)
67. Gibtan A, Getahun A, Mengistou S. Effect of stocking density on the growth performance and yield of Nile tilapia [*Oreochromis niloticus*(L., 1758)] in a cage culture system in Lake Kuriftu, Ethiopia. *Aquaculture Research*. 2008;39(13):1450-1460. [doi:10.1111/j.1365-2109.2008.02021.x](https://doi.org/10.1111/j.1365-2109.2008.02021.x)
68. Antony J, Balasubramanian CP, Balamurugan J, Sandeep KP, Biju IF, Vijayan KK. Optimisation of nursery rearing for megalopa of giant mud crab *Scylla serrata* (Forsk., 1775). *Indian Journal of Fisheries*. 2019;66(1):43-50. [doi:10.21077/ijf.2019.66.1.80293-06](https://doi.org/10.21077/ijf.2019.66.1.80293-06)
69. Hamasaki K, Suprayudi MA, Takeuchi T. Mass mortality during metamorphosis to megalops in the seed production of mud crab *Scylla serrata* (Crustacea, Decapoda, Portunidae). *Fisheries Sci*. 2002;68(6):1226-1232. [doi:10.1046/j.1444-2906.2002.00559.x](https://doi.org/10.1046/j.1444-2906.2002.00559.x)
70. Mann DL, Asakawa T, Kelly B, Lindsay T, Paterson B. Stocking density and artificial habitat influence stock structure and yield from intensive nursery systems for mud crabs *Scylla serrata* (Forsskål 1775). *Aquaculture Res*. 2007;38(14):1580-1587. [doi:10.1111/j.1365-2109.2006.01626.x](https://doi.org/10.1111/j.1365-2109.2006.01626.x)